Outlining the maturational pathway of mitochondrial short non-coding RNAs using in silico and in vitro approaches Alessandro Formaggioni*, Federico Plazzi, Marco Passamonti More at: University of Bologna. Department of Biological, Geological and Environmental Sciences. Bologna. alessand.formaggion2@unibo.it X ale97_28 References 1. Passamonti et al.; 2020; Sci. Rep.;10(1):8219 2. Farberov et al.; 2023; *iScience*, 26(10):107723 3. Pozzi et al.; 2020; *GBE*; 14(2):evac023 In silico analysis using CLIP-seq libraries Intro The Small Mitochondrial Highly Transcribed RNAs (smithRNAs) are a IP of target RNA-protein Enriched pool of complexes protein RNAs new class of small interfering RNAs transcribed in the mt-genome regulating nuclear targets. Their effects were in vivo validated in the Manila clam, Ruditapes philippinarum. Moreover, in silico analysis predicted that smitRNAs are widespread across metazoans¹. The biogenesis of smithRNAs is not well defined yet. We analyzed publicly available CLIP-seq data to detect which proteins may interact with In vertebrates: DROSHA+DGCR8 and AGO2 participate respectively to the first and last smithRNAs. Moreover, synthetized smithRNA were pulled down with step of the miRNA pathway. Analyzing CLIP-seq libraries of these proteins, all three R. philippinarum proteins.

proteins were predicted to interact with mt-RNAs (Fig. 2). The most enriched mt-RNAs are

In vitro analysis using RNA-protein pull-down

Biotin

Pull-down analysis with biotinylated smithRNAs (122nca and 145t), one nuclear miRNA (let7), and their preliminary structures (pre_), to identify which proteins interact with these RNAs in R. philippinarum. Streptavidin



proteins detected by the mass spectrometer in each sample. Preliminary RNAs (pre_122nca, pre_145t, pre_let7) cluster together, as well as two mature small RNAs (122nca, let7).

We selected proteins that were detected at two-fold higher levels in a pulled-down sample compared to the control sample. These proteins are significantly associated with specific GO terms: let-7 and 145t interact with isomerases (cycliophilin and rotamase).

all located in tRNA regions (tRNA-Val, tRNA-Phe, tRNA-Met), this signal is conserved in homologous regions in M. musculus.

	DRO	SHA	DG	CR8	AC	602		AGO2	
Cell Line:	HepG2	K562	HepG2	K562	HCT 116	HCT 116	n	eurons	>
tRNA-Val (1614-1651)-	2.8*	4.98*	2.27*	2.12*	1.65*	1.91*	tRNA-Val (1037-1072)-	1.62*	
tRNA-Phe (614-645)-	2.62*	2.79*	1.48*	1.31*	1.85*	3*	tRNA-Phe (33-64)-	1.95*	
tRNA-Met (4419-4461)-	1.62*	2.62*	0.35	1.03*	1.62*	4*	tRNA-Met (3863-3905)-	2.91*	
tRNA-Pro (15957-15989)-	0.3	3.22*	0.4	1.32*	3.16*	1.97*	tRNA-Pro (15356-15387)-	2.2*	
tRNA-Gly (10008-10054)-	1.28*	3.84*	0.39	1.67*	2.27*	1.97*	tRNA-Gly (9408-9454)-	0.39	
d-loop (327-396)-	0.84*	3.13*	-0.43	0.59	1.87*	2.72*	d-loop (16296-16365)-	1.39	
s-rRNA (1525-1542)-	1.45*	2.44*	0.16	-0.16	1.39*	2.08*	s-rRNA (968-985)-	0.78*	logEC
s-rRNA (1390-1500)-	1.11*	1.66*	-0.52	0.6*	-0.03	2.45*	s-rRNA(803-943)-	1.48*	
s-rRNA (709-769)-	0.67*	1.42*	-0.13	0.57*	-0.49	2.71*	s-rRNA(122-182)-	1.27*	2.8 0.0
tRNA-Tyr (5828-5859)-	-0.03	2.61*	-0.17	0.77	2.48*	1.22*	tRNA-Tyr (5262-5293)-	1.39	-2
s-rRNA (1272-1377)-	0.79*	1.57*	-0.62	0.01	-0.65	2.19*	s-rRNA (685-790)-	1.5*	
NADH6 (14644-14501)-	0.39	2.06*	-0.02	0	1.92*	2.02*	NADH6 (13990-14047)-	-1.33	
COX1 (6909-6942)-	-0.3	0.88*	-0.94	-0.13	-0.2	2.36*	COX1 (6333–6366)-	2.62*	
COX1 (6508-6527)-	-0.48	0.71*	-1.76	-0.21	0.55	2.22*	COX1 (5932-5951)-	1.83*	
tRNA-Thr (15920-15953)-	-0.84	0.01	-2.56	-1.72	2.95*	1.42	tRNA-Thr (15322-15355)-	1.16	
tRNA-Glu (14677-14708)-	-0.58	0.88	-0.24	0.14	1.36	2.02*	tRNA-Glu (14704-14735)-	-0.39	

2.5

0.0

-2.5

	Description	let7	145t	122nca
Tab. 1A : : GO terms enriched in each small	Cycliophilin type peptidyl-prolyl cis-trans isomerase	5.03E-07	2.00E-06	Х
RNA with the respective	RNA recognition motif	X	8.46E-08	Х
p-value. 'X' when the	mRNA processing	Х	Х	1.52E-02
significant.	Rotamase	4.51E-06	9.55E-06	Х

Preliminary structures were more associated with spliceosome activity, in particular in pre_145t and pre_122nca:

	Description	pre_let7	pre_145t	pre_122nca
Tab. 1B: : GO terms	RNA recognition motif	3.82E-07	6.60E-07	4.34E+07
enriched in each small	mRNA splicing	Х	7.05-E07	5.58E-07
RNA with the respective p-value. 'X' when the	Nucleotide-binding alpha-beta plait domain	2.39E-06	3.89E-07	2.52E-07
significant.	Spliceosome	Х	1.04-E07	4.61E-07
-				

Many GO terms were associated with splicesome activity (cycliophylin, mRNA processing, mRNA splicing, splicing). miRNAs were reported to have a role in the modulation of splicing²

Conclusions

In silico and in vitro data suggest that smithRNAs could interact with proteins belonging to the microRNA pathway. In particular, we showed that AGO2 preferentially interacts with tRNAs (where most smithRNAs are located^{1,3}) in the mt-genome of three different species. Mt-tRNAs were also predicted to interact with the Microprocessor complex

Figure 2: all the mt-regions that showed a logFC (log₂(n° mapping reads IP library / n° mapping reads control library)) > 2 in at least one of the samples were reported in the figure. Boxes are colored in shades of reds when logFC > 0 and in shades of blue when logFC < 0. Significant logFC values are marked with *.

In C. elegans: we analyzed the RIP-seq libraries of 4 argonaute proteins (CSR1, WAGO1, HRDE1, ALG1). Only CSR1 was predicted to ineract with mt-RNAs (Tab. 2A), which are located in coding or unassigned regions (UR).

	Region	logFC	logCPM	P-value	
	UR:13315-13358	3.87	11.90	1.94E-04	
	CYTB:4882-4926	3.79	8.28	2.67E-05	
	CYTB:5250-5291	2.61	6.26	6.00E-04	
Tab. 2A : significantly DE mt-	CYTB:5293-5340	3.57	6.24	5.13E-05	
small RNAs in CSR-1 IP	NADH5:12350-12390	1.94	4.56	1.38E-02	
libraries vs control libraries.	NADH1:1794-1843	3.39	4.59	8.84E-05	
					1

In D. melanogaster: we analyzed RIP-seq libraries of 5 argonaute proteins (AGO1-3, PIWI and Aubergine). Singificanlty enriched mt-regions were found only in the AGO2-IP libraries. Each region is located in a different tRNA. tRNA-Met is by far the most

enriched	(Tab.	2B).	
	(100)		

Region	logFC	logCPM	P-value	
tRNA-Met:167-187	9.95	9.21	2.20E-09	
tRNA-Leu1:3009-3032	1.22	5.30	4.30E-03	
tRNA-Ala:5982-6001	2.00	5.05	1.59E-04	



